

DAFTAR PUSTAKA

- Aisyah, S. N., Sulastri, S., Retmi, R., Yani, R. H., Syafriani, E., Syukriani, L., Fatchiyah, F., Bakhtiar, A., & Jamsari, J. (2017). Suppression of *Colletotrichum gloeosporioides* by Indigenous *Phyllobacterium* and its Compatibility with Rhizobacteria. *Asian Journal of Plant Pathology*, *11*(3), 139–147. <https://doi.org/10.3923/ajppaj.2017.139.147>.
- Akbar, R., Robert, P. A., Pavlović, M., Jeliakov, J. R., & Snapkov, I. (2023). Konstruksi dan Ekspresi Protein Rekombinan Wee1 pada Plasmid pET28a. *Jurnal Ilmu Dasar*, *24*(2), 151-158.
- Al Doghaither, H., & Gull, M. (2019). Plasmids as genetic tools and their applications in ecology and evolution. In *Plasmid*. IntechOpen. <https://doi.org/10.5772/intechopen.81271>.
- Alekseeva, I. V., & Kuznetsov, N. A. (2023). Historical aspects of restriction endonucleases as intelligent scissors for genetic engineering. *Fermentation*, *9*(10), 874, <https://doi.org/10.3390/fermentation9100874>.
- Aliya, S. L. (2025). *Fusi Gen ansB Dengan Promoter Sintetik pSSPM3 ke Dalam Vektor Ekspresi pET-28a+ Untuk Pengembangan Terapi Target Acute Lymphoblastic Leukemia* [Skripsi, Universitas Andalas]. Universitas Andalas.
- Alper, H., Fischer, C., Nevoigt, E., & Stephanopoulos, G. (2005). Tuning genetic control through promoter engineering. *Nature Biotechnology*, *23*, 612–616. <https://doi.org/10.1038/nbt1082>.
- Amalia, A. R. (2025). *Optimasi ekspresi enzim rekombinan Pfu DNA polimerase pada sistem ekspresi Escherichia coli BL21 dengan agen penginduksi isopropil-β-d-tiogalaktopiranosida (IPTG)* [Skripsi, UIN Sunan Kalijaga Yogyakarta]. UIN Sunan Kalijaga Yogyakarta.
- Arslan, M., Tezcan, E., Camcı, H., & Avcı, M. K. (2021). Effect of DNA concentration on band intensity and resolution in agarose gel electrophoresis. *Van Sağlık Bilimleri Dergisi*, *14*(3), 326-333.
- Aulia, S. L., Suwignyo, R. A., & Hasmeda, M. (2021). Optimasi Suhu Annealing untuk Amplifikasi DNA Padi Hasil Persilangan Varietas Tahan Terendam dengan Metode Polymerase Chain Reaction. *Sainmatika: Jurnal Ilmiah Matematika dan Ilmu Pengetahuan Alam*, *18*(1), 44-54.
- Avendano, K. A., Anguiano, M., Lopez, C. E., Montanez, L. E., Sifuentes, L., & Balagurusamy, N. (2016). Microbial enzymes applications in food processing. *Agro Food Industry Hi-Tech*, *27*(4), 63-67.

- Avramis, V. I., & Tiwari, P. N. (2006). Asparaginase (native and pegylated) in the treatment of acute lymphoblastic leukemia. *International Journal of Nanomedicine*, 1(3), 241-254.
- Baeshen, N. A., Baeshen, M. N., Sheikh, A., Bora, R. S., Ahmed, M. M. M., Ramadan, H. A., & Redwan, E. M. (2014). Cell factories for insulin production. *Microbial cell factories*, 13(1), 141. <https://doi.org/10.1186/s12934-014-0141-0>.
- Bansal, M., Kumar, A., & Yella, V. R. (2014). Role of DNA sequence based structural features of promoters in transcription initiation and gene expression. *Current opinion in structural biology*, 25, 77-85. <https://doi.org/10.1016/j.sbi.2014.01.007>.
- Bartnik, K., Barth, A., Pilo-Pais, M., Crevenna, A. H., Liedl, T., & Lamb, D. C. (2019). A DNA Origami Platform for Single-Pair Förster Resonance Energy Transfer Investigation of DNA–DNA Interactions and Ligation. *Journal of the American Chemical Society*, 142(2), 815-825. <https://doi.org/10.1021/jacs.9b10073>.
- Basiri, H., Akbari, N., Azizpour, M., Hosseini, S. D., Behrozikhah, A. M., & Eskandari, S. (2013). Amplification, cloning and expression of *Brucella melitensis* bp26 gene (OMP28) isolated from Markazi province (Iran) and purification of Bp26 Protein. *Archives of Razi Institute*, 68(2), 111-116.
- Batool, T., Makky, E. A., Jalal, M., & Yusoff, M. M. (2016). A Comprehensive Review on L-Asparaginase and Its Applications. *Applied biochemistry and biotechnology*, 178(5), 900–923. <https://doi.org/10.1007/s12010-015-1917-3>.
- Bernadus, Z. G., Fatimawali, F., & Kolondam, B. (2019). Transformasi Plasmid yang Mengandung Gen *Merb* pada *Escherichia coli* B121 (DE3). *PHARMACON*, 8(1), 196-202.
- Beutel, B. A., & Record Jr, M. T. (1990). *E. coli* promoter spacer regions contain nonrandom sequences which correlate to spacer length. *Nucleic acids research*, 18(12), 3597-3603. <https://doi.org/10.1093/nar/18.12.3597>.
- Blazeck, J., Garg, R., Reed, B., & Alper, H. S. (2012). Controlling promoter strength and regulation in *Saccharomyces cerevisiae* using synthetic hybrid promoters. *Biotechnology and bioengineering*, 109(11), 2884-2895. <https://doi.org/10.1002/bit.24563>.
- Blount, B. A., Weenink, T., Vasylechko, S., & Ellis, T. (2012). Rational diversification of a promoter providing fine-tuned expression and orthogonal regulation for synthetic biology. *PloS one*, 7(3), e33279. <https://doi.org/10.1371/journal.pone.0033279>.

- Bolivar, F., & Backman, K. (1979). Plasmids of *Escherichia coli* as cloning vectors. In *Methods in enzymology* (Vol. 68, pp. 245-267). Academic Press. [https://doi.org/10.1016/S0076-6879\(79\)68018-3](https://doi.org/10.1016/S0076-6879(79)68018-3).
- Browning, D. F., & Busby, S. J. W. (2004). The regulation of bacterial transcription initiation. *Nature Reviews Microbiology*, 2, 57–65. <https://doi.org/10.1038/nrmicro787>.
- Darvishi, F., Jahanafrooz, Z., & Mokhtarzadeh, A. (2022). Microbial L-asparaginase as a promising enzyme for treatment of various cancers. *Applied microbiology and biotechnology*, 106(17), 5335–5347. <https://doi.org/10.1007/s00253-022-12150-2>.
- De Boer, H. A., Comstock, L. J., & Vasser, M. (1983). The tac promoter: a functional hybrid derived from the trp and lac promoters. *PNAS*, 80(1), 21–25. <https://doi.org/10.1073/pnas.80.1.21>.
- Deal, C., De Wannemaeker, L., & De Mey, M. (2024). Towards a rational approach to promoter engineering: understanding the complexity of transcription initiation in prokaryotes. *FEMS Microbiology Reviews*, 48(2), fuae004. <https://doi.org/10.1093/femsre/fuae004>.
- Debode, F., Marien, A., Janssen, É., Bragard, C., & Berben, G. (2017). The influence of amplicon length on real-time PCR results. *Biotechnologie, Agronomie, Société et Environnement*, 21(1), 3–11. <https://doi.org/10.25518/1780-4507.15096>.
- Deng, J., Wu, Y., Zheng, Z., Chen, N., Luo, X., Tang, H., & Keasling, J. D. (2021). A synthetic promoter system for well-controlled protein expression with different carbon sources in *Saccharomyces cerevisiae*. *Microbial cell factories*, 20(1), 202. <https://doi.org/10.1186/s12934-021-01692-9>.
- Dubendorf, J. W., & Studier, F. W. (1991). Controlling basal expression in an inducible T7 expression system by blocking the target T7 promoter with lac repressor. *Journal of molecular biology*, 219(1), 45-59. [https://doi.org/10.1016/0022-2836\(91\)90801-E](https://doi.org/10.1016/0022-2836(91)90801-E).
- Egler, R. A., Ahuja, S. P., & Matloub, Y. (2016). L-Asparaginase in the treatment of patients with acute lymphoblastic leukemia. *Journal of Pharmacology & Pharmacotherapeutics*, 7(2), 62-71. <https://doi.org/10.4103/0976-500X.182882>.
- Feng, Y., Xie, Z., Jiang, X., Li, Z., Shen, Y., Wang, B., & Liu, J. (2018). The applications of promoter-gene-engineered biosensors. *Sensors*, 18(9), 2823. <https://doi.org/10.3390/s18092823>.
- Gourse, R. L., Ross, W., & Gaal, T. (2000). UPs and downs in bacterial transcription initiation: the role of the alpha subunit of RNA polymerase in promoter

- recognition. *Molecular microbiology*, 37(4), 687-695.
<https://doi.org/10.1046/j.1365-2958.2000.02096.x>.
- Green, M. R., & Sambrook, J. (2020). Cloning in plasmid vectors: blunt-end cloning. *Cold Spring Harbor Protocols*, 2020(11), pdb-prot101246.
<https://doi.org/10.1101/pdb.prot101246>.
- Green, M.R. and Sambrook, J. (2020). 'The Inoue Method for Preparation and Transformation of Competent *Escherichia coli*: "Ultracompetent" Cells', *Cold Spring Harbor Protocols*, (6), p. pdb.prot101196.
<https://doi.org/10.1101/pdb.prot101196>.
- Gummesson, B., Lovmar, M., & Nyström, T. (2013). A proximal promoter element required for positive transcriptional control by guanosine tetraphosphate and DksA protein during the stringent response. *Journal of Biological Chemistry*, 288(29), 21055-21064.
<https://doi.org/10.1074/jbc.M113.479998>.
- Hahn, M. B. (2025). Rapid quantitative analysis of double-stranded plasmid DNA with capillary gel electrophoresis for applications in quality control and radiation research. *Scientific Reports*, 15(1), 1068.
- Hansur, L., Ugi, D., & Hambali, H. (2019). Uji kepekaan bakteri asam laktat kandidat probiotik terhadap antibiotik kanamisin, oleandomisin, dan polimiksin B. *eJKI*, 7(1).
- Harimadi, K. J., Milka, M., El Kiyat, W., & Budijanto, S. (2018). Potensi pemanfaatan asparaginase untuk mengurangi kadar akrilamida pada keripik kentang dan singkong. *Jurnal Pangan*, 27(1), 67-78.
- Hikmatyar, M. F., & Royani, J. I. (2015). Isolasi dan amplifikasi DNA keladi tikus (*Thyponium flagelliform*) untuk identifikasi keragaman genetik. *Jurnal Bioteknologi & Biosains Indonesia (JBBI)*, 2(2), 42-48.
- Indriarini, D., Rukmana, A., & Yasmon, A. (2018). Cloning and expression of MCE1A gene from *Mycobacterium tuberculosis* Beijing and H37RV strain for vaccine candidate development. *African journal of infectious diseases*, 12(1S), 127-132.
- Ishihama, A. (1997). Promoter selectivity control of RNA polymerase. In *Mechanisms of Transcription* (pp. 53-70). Berlin, Heidelberg: Springer Berlin Heidelberg.
- Ito, Y., Asayama, M., & Shirai, M. (2003). Light-responsive psbA transcription requires the -35 hexamer in the promoter and its proximal upstream element, UPE, in cyanobacteria. *Bioscience, biotechnology, and biochemistry*, 67(6), 1382-1390.
- Jana, A., Biswas, S., Ghosh, R., & Modak, R. (2024). Recent advances in L-Asparaginase enzyme production and formulation development for

- acrylamide reduction during food processing. *Food chemistry: X*, 25, 102055. <https://doi.org/10.1016/j.fochx.2024.102055>.
- Jannah, M. (2023). Optimalisasi Kondisi PCR Untuk Amplifikasi Sekuen Gen HBB. *ORYZA: Jurnal Pendidikan Biologi*, 12(1), 36-42. <https://doi.org/10.33627/oz.v12i1.1057>.
- Jia, R., Wan, X., Geng, X., Xue, D., Xie, Z., & Chen, C. (2021). Microbial L-asparaginase for Application in Acrylamide Mitigation from Food: Current Research Status and Future Perspectives. *Microorganisms*, 9(8), 1659. <https://doi.org/10.3390/microorganisms9081659>.
- Kiryama, Y., Kubota, M., Takimoto, T., Kitoh, T., Tanizawa, A., Akiyama, Y., & Mikawa, H. (1989). Biochemical characterization of L-Asparaginase in mammals. *Leukemia Research*, 13(6), 451-457.
- Krishnapura, P. R., Belur, P. D., & Subramanya, S. (2015). A critical review on properties and applications of microbial l-asparaginases. *Critical reviews in microbiology*, 42(5), 720-737. <https://doi.org/10.3109/1040841X.2014.946555>.
- Langden, S. S., Budiharjo, A., & Kusharyoto, W. (2017). Transformasi dan Kloning Plasmid PJ804:77539 Pada *E. coli* Top'10. *Jurnal Akademika Biologi*, 6(1), 65-70.
- Li, J., Wang, Y., Zhang, Y., Ye, Z., & Yang, Y. (2019). Characterization of tissue-specific gene promoters in ramie (*Boehmeria nivea* L.). *BMC Plant Biology*, 19, 338. <https://doi.org/10.1186/s12870-019-1880-z>.
- Li, L., Li, H., Tian, Q., Ge, B., Xu, X., Chi, Y., ... & Zhou, Y. (2022). Expression and purification of soluble recombinant β -lactamases using *Escherichia coli* as expression host and pET-28a as cloning vector. *Microbial Cell Factories*, 21(1), 244. <https://doi.org/10.1186/s12934-022-01885-w>.
- Liu, J., Chang, W., Pan, L., Liu, X., Su, L., Zhang, W., Li, Q. & Zheng, Y. (2018). An Improved Method of Preparing High Efficiency Transformation *Escherichia coli* with Both Plasmids and Larger DNA Fragments, *Indian Journal of Microbiology*, 58(4), 448-456. <https://doi.org/10.1007/s12088-018-0735-7>.
- Liu, Z. Q., & Yang, P. C. (2012). Construction of pET-32 α (+) vector for protein expression and purification. *North American journal of medical sciences*, 4(12), 651.
- Loch, J. I., & Jaskolski, M. (2021). Structural and biophysical aspects of L-asparaginases: a growing family with amazing diversity. *IUCrJ*, 8(4), 514-531. <https://doi.org/10.1107/S2052252521008300>.
- Lopes, A. M., Oliveira-Nascimento, L., Ribeiro, A., Tairum, C. A., Breyer, C. A., Oliveira, M. A., & Pessoa, A. (2017). Therapeutic L-Asparaginase:

- Upstream, downstream and beyond. *Critical Reviews in Biotechnology*, 37(1), 82-99. <https://doi.org/10.3109/07388551.2016.1149285>.
- Lubkowski, J., et al. (2020). Structural and biochemical studies of therapeutic enzymes. *Advances in Protein Chemistry and Structural Biology*, 120, 1 – 35. <https://doi.org/10.1111/febs.16042>.
- Makino, T., Skretas, G., & Georgiou, G. (2011). Strain engineering for improved expression of recombinant proteins. *Protein Expression and Purification*, 79(3), 166–171. <https://doi.org/10.1186/1475-2859-10-32>.
- Maksum, I. P., Budiantoro, O., Hasan, K., Soemitro, S., & Subroto, T. (2017). Pemurnian pretrombin-2pH hasil ko-ekspresi chaperone pada *Escherichia coli* ER2566 menggunakan sistem IMPACT. *Chimica et Natura Acta*, 5(2), 57-64. <https://doi.org/10.24198/cna.v5.n2.14605>.
- Mazlan, A. H., Muhamad Najib, M. H. A., Hazizul Hassan, M., Mohd Hatta, F. H., & Mohd Yusoff, R. (2024). Effect of DNA template concentration on standard polymerase chain reaction. *International Journal of Pharmaceutics, Nutraceuticals and Cosmetic Science (IJPNaCS)*, 7(1), 1-11.
- Mierendorf, R. C., Morris, B. B., Hammer, B., & Novy, R. E. (1998). Expression and purification of recombinant proteins using the pET system. In *Molecular diagnosis of infectious diseases* (pp. 257-292). Totowa, NJ: Humana Press.
- Mirayanti, N. L. W. Y., Pharmawati, M., & Mahardika, I. G. N. K. (2022). Produksi Rekombinan *Bovine Lactoferrin* pada Sistem Ekspresi *Escherichia coli*. *Jurnal Veteriner*, 23(1), 105-111.
- Mollah, A., Ashan, M. A., & Khatimah, A. H. (2022). Uji kualitas dan kuantitas DNA porang (*Amorphophallus Muelleri Blume*) pada beberapa kawasan di sulawesi *Jurnal Agritechno*, 15 (01), 1–7.
- Mumm, J. P., Friedman, L. J., & Gelles, J. (2020). Mechanism of upstream promoter element stimulation of transcription at a ribosomal RNA promoter determined by single-molecule imaging. *BioRxiv*, 2020-02.
- Nouemssi, S. B., Ghribi, M., Beauchemin, R., Meddeb-Mouelhi, F., Germain, H., & Desgagné-Penix, I. (2020). *Rapid and efficient colony-PCR for high throughput screening of genetically transformed Chlamydomonas reinhardtii*. *Life*, 10(9), 186. <https://doi.org/10.3390/life10090186>.
- Oza, V. P., Parmar, P. P., Kumar, S., & Subramanian, R. B. (2011). Anticancer properties of highly purified L-Asparaginase from *Withania somnifera* L. *Journal of Biotechnology*, 152(4), 131-136.
- Paradiman, A. Z., Tahir, M. M., & Dirpan, A. (2024). Formation of acrylamide compounds in food products from maillard reactions: A review article. In

BIO Web of Conferences, 96, 01030.
<https://doi.org/10.1051/bioconf/20249601030>.

Pedreschi, F., et al. (2008). The effect of asparaginase on acrylamide formation in French fries. *Food Chemistry*, 109(2), 386–392.
<https://doi.org/10.1016/j.foodchem.2007.12.057>.

Pokrovskaya, M. V., Pokrovsky, V. S., Aleksandrova, S. S., Sokolov, N. N., & Zhdanov, D. D. (2022). Molecular analysis of L-asparaginases for clarification of the mechanism of action and optimization of pharmacological functions. *Pharmaceutics*, 14(3), 599.
<https://doi.org/10.3390/pharmaceutics14030599>.

Polosoro, A., Enggarini, W., Kusumanegara, K., Hadiarto, T., Miftahudin, M., & Supena, E. D. J. (2024). Optimasi Ekstraksi RNA dan Teknik Kloning: Studi Kasus Kloning Gen Heading Date 3a pada Kelapa Sawit. *Vegetalika*, 13(2), 196-208.

Pratiwi, R. D. (2019). Optimasi ekspresi human Epidermal Growth Factor (h-EGF) rekombinan dalam *Escherichia coli* BL21 (DE3) dengan variasi media dan konsentrasi penginduksi. *Chimica et Natura Acta*, 7(2), 91-97.

Pui, C. H., & Evans, W. E. (2006). Treatment of acute lymphoblastic leukemia. *New England Journal of Medicine*, 354(2), 166-178.
<https://doi.org/10.1056/NEJMra052603>.

Qi, Z., Han, J., Zhang, H., Wang, Y., & Liu, Y. (2020). A cambium-specific promoter from ramie allows targeted gene expression in vascular tissues of herbaceous plants. *Plant Molecular Biology Reporter*, 38, 394–405.
<https://doi.org/10.1007/s11105-020-01185-2>.

Rodiansyah, A., Tan, M. I., & Nugrahapraja, H. (2022). Construction, cloning, and overexpression of staphylococcal enterotoxin B gene synthetic (SEBSyn) in pET-28a (+): pre-development bacterial-toxin therapy for cancer. In *7th International Conference on Biological Science (ICBS 2021)* (pp. 464-470). Atlantis Press.

Rosano, G. L., & Ceccarelli, E. A. (2014). Recombinant protein expression in *Escherichia coli*: advances and challenges. *Frontiers in microbiology*, 5, 172. <https://doi.org/10.3389/fmicb.2014.00172>.

Ross, W., Gosink, K. K., Salomon, J., Igarashi, K., Zou, C., Ishihama, A., ... & Gourse, R. L. (1993). A third recognition element in bacterial promoters: DNA binding by the α subunit of RNA polymerase. *Science*, 262(5138), 1407-1413. <https://doi.org/10.1126/science.8248780>.

Ruff, E. F., Record Jr, M. T., & Artsimovitch, I. (2015). Initial events in bacterial transcription initiation. *Biomolecules*, 5(2), 1035-1062.
<https://doi.org/10.3390/biom5021035>.

- Sanches, M., Krauchenco, S., & Polikarpov, I. (2007). Structure, substrate complexation and reaction mechanism of bacterial asparaginases. *Current Chemical Biology*, 1(1), 75-86.
- Septiasari, N. P. S., Junitha, I. K., & Wirasiti, N. N. (2023). Optimasi digesti enzim restriksi untuk deteksi mutasi daerah D-loop DNA mitokondria dengan metode PCR-RFLP. *Jurnal Biologi Udayana*, 27(1), 65-72. <https://doi.org/10.24843/JBIOUNUD.2023.v27.i01.p07>.
- Setiani, N. A., Baroroh, U., Handayani, A. P., Chairunnisa, N., & Mardiah, I. (2022). Konstruksi Plasmid Rekombinan pET-28a-Nanobodi Secara In Silico. In *Prosiding Seminar Nasional Diseminasi Hasil Penelitian Program Studi S1 Farmasi* (Vol. 2, No. 1)..
- Setyawati, R., & Zubaidah, S. (2021). Optimasi konsentrasi primer dan suhu annealing dalam mendeteksi gen *leptin* pada sapi peranakan ongole (PO) menggunakan polymerase chain reaction (PCR). *Indonesian Journal of Laboratory*, 4(1), 36-40.
- Shrivastava, A., Khan, A. A., Khurshid, M., Kalam, M. A., Jain, S. K., & Singhal, P. K. (2016). Recent developments in L-Asparaginase discovery and its potential as anticancer agent. *Critical Reviews in Oncology/Hematology*, 100, 1-10. <https://doi.org/10.1016/j.critrevonc.2015.01.002>.
- Silva-Santos, A. R., Alves, C. P., Prazeres, D. M. F., & Azevedo, A. M. (2016). Separation of plasmid DNA topoisomers by multimodal chromatography. *Analytical biochemistry*, 503, 68-70. <https://doi.org/10.1016/j.ab.2016.02.018>.
- Susanto, A. H., Pramono, H., & Lestari, P. (2009). Construction of Soil Metagenomic Library to Obtain Recombinant Clones with an Indigenous Lipase Activity. *Biota: Jurnal Ilmiah Ilmu-Ilmu Hayati*, 150-155.
- Sztiller-Sikorska, M., Heyduk, E., & Heyduk, T. (2011). Promoter spacer DNA plays an active role in integrating the functional consequences of RNA polymerase contacts with -10 and -35 promoter elements. *Biophysical chemistry*, 159(1), 73-81. <https://doi.org/10.1016/j.bpc.2011.06.005>.
- Tabor, S. (1990). Expression using the T7 RNA polymerase/promoter system. *Current protocols in molecular biology*, 11(1), 16-2. <https://doi.org/10.1002/0471142727.mb1602s11>.
- Terpe, K. (2006). Overview of bacterial expression systems for heterologous protein production: from molecular and biochemical fundamentals to commercial systems. *Applied microbiology and biotechnology*, 72(2), 211-222. <https://doi.org/10.1007/s00253-006-0465-8>.
- Thermo Fisher Scientific. (2024). *PCR component considerations*. Thermo Fisher Scientific. Diakses dari <https://www.thermofisher.com/id/en/home/life->

science/cloning/cloning-learning-center/invitrogen-school-of-molecular-biology/pcr-education/pcr-reagents-enzymes/pcr-component-considerations.html

- Ullah, A., Bashir, A., Rehman, B., Naeem, W., & Shah, S. Z. (2023). Optimization of colony polymerase chain reaction for the 16S rRNA of different strains of *Escherichia coli*. *Innovare Journal of Life Sciences*, 11, 32–35.
- Versmessen, N., Van Simaey, L., Negash, A. A., Vandekerckhove, M., Hulpiau, P., Vanechoutte, M., & Cools, P. (2024). Comparison of DeNovix, NanoDrop and Qubit for DNA quantification and impurity detection of bacterial DNA extracts. *PLoS one*, 19(6), e0305650. <https://doi.org/10.1371/journal.pone.0305650>.
- Wang, Y., Shi, H., Sun, D., Zheng, X., He, G., & Yang, Y. (2016). A vascular-specific promoter from ramie (*Boehmeria nivea*) drives PVX gene expression in *Arabidopsis*. *International Journal of Molecular Sciences*, 17(7), 1109. <https://doi.org/10.3390/ijms17071109>.
- Wang, Z., Jin, L., Yuan, Z., Węgrzyn, G., & Węgrzyn, A. (2009). Classification of plasmid vectors using replication origin, selection marker and promoter as criteria. *Plasmid*, 61(1), 47-51. <https://doi.org/10.1016/j.plasmid.2008.10.002>.
- Winkelman, J. T., Vvedenskaya, I. O., Zhang, Y., Zhang, Y., Bird, J. G., Taylor, D. M., & Nickels, B. E. (2016). Multiplexed protein-DNA cross-linking: Scrunching in transcription start site selection. *Science*, 351(6277), 1090-1093. <https://doi.org/10.1126/science.aad7365>.
- World Health Organization. (2011). *Safety evaluation of certain contaminants in food: acrylamide (63rd)*. Roma: World health organization.
- Xu, F., Oruna-Concha, M. J., & Elmore, J. S. (2016). The use of asparaginase to reduce acrylamide levels in cooked food. *Food chemistry*, 210, 163-171. <https://doi.org/10.1016/j.foodchem.2016.04.095>.
- Zainuddin, M., Pringgenies, D., Radjasa, O. K., Haeruddin, H., Sabdaningsih, A., & Herawati, V. E. (2022). Optimasi Kondisi Inkubasi Kultur (Suhu dan Agitasi) Terhadap Pertumbuhan dan Aktivitas Protease Ekstraseluler Bakteri *Bacillus firmus* Asosiasi Sponge (Porifera: Demospongiae) dari Nusa Lembongan Bali Indonesia. *Journal of Marine Research*, 11(3), 547-556.
- Zhivagui, M., Ng, A. W. T., Ardin, M., Churchwell, M. I., Pandey, M., Renard, C., et al. (2019). Experimental and pan-cancer genome analyses reveal widespread contribution of acrylamide exposure to carcinogenesis in humans. *Genome Research*, 29(4), 521–531. <https://doi.org/10.1101/gr.243933.118>.